

Synthesis of Peptides containing the β -substituted Aminoethane Sulfonamide or Sulfonamide Transition-state Isostere derived from Amino Acids

Wilna J. Moree, Gijs A. van der Marel and Rob M.J. Liskamp*

Gorlaeus Laboratories, University of Leiden, P.O. Box 9502, 2300 RA Leiden, The Netherlands

Abstract: α -amino acids can be converted to homochiral β -substituted aminoethane sulfonamide or sulfonamide transition-state isosteres, which can be incorporated into peptides. These transition-state analogues e.g. the sulfonamide isostere of Phe-Phe will be used for the generation of catalytic antibodies as well as for the development of protease inhibitors.

Transition-state analogues of the amide bond hydrolysis are important both for the design of enzyme inhibitors¹ and the development of catalytic antibodies². Therefore numerous transition-state isosteres of the amide bond hydrolysis have been described¹⁻³. Sulfonamides⁴ and sulfonamides^{4,5} have received relatively little attention, in spite of their good resemblance to the amide bond hydrolysis⁶. In a recent communication⁶ we reported the synthesis of peptides incorporating the β -aminoethane sulfonamide or sulfonamide transition-state analogues (scheme 1, 1 and 2). These were proposed to mimic the amide bond hydrolysis between glycine and any other amino acid or peptide. In addition, we found that upon α -alkylation of sulfonamides (2) analogues of other amino acids than glycine (3) became accessible. This enabled us to prepare sulfonamides (figure 1b) mimicking amide bond hydrolysis of the p17/p24 cleavage site Phe-Pro (figure 1a) in the gag-pol precursor protein of HIV⁷, thus giving rise to a new type of potential HIV protease inhibitors⁸.

In contrast to α -aminosulfonamides or sulfonamides^{4,9} the corresponding β -aminoethane derivatives 1 and 2 are stable. Moreover the presence of the additional β -carbon atom offers the opportunity to study the influence of functionalization at this position (figure 1c) as compared to functionalization at the α -position. Based on the method of Higashiura and Ienaga¹⁰ we developed a method for the synthesis of homochiral β -substituted sulfonamide (4) and sulfonamide (5) transition-state isosteres incorporated in peptides. Since the isosteres can be derived from naturally occurring α -amino acids in principle all amino acids can be used as a starting material for the preparation of these potentially interesting compounds (scheme 1). This adds to the scope of this procedure (*vide infra*).

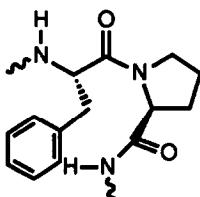


figure 1a

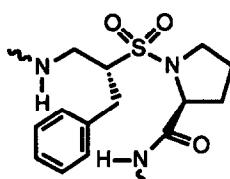


figure 1b

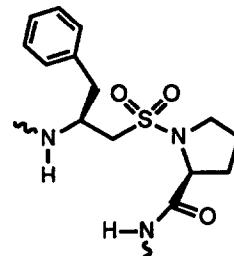
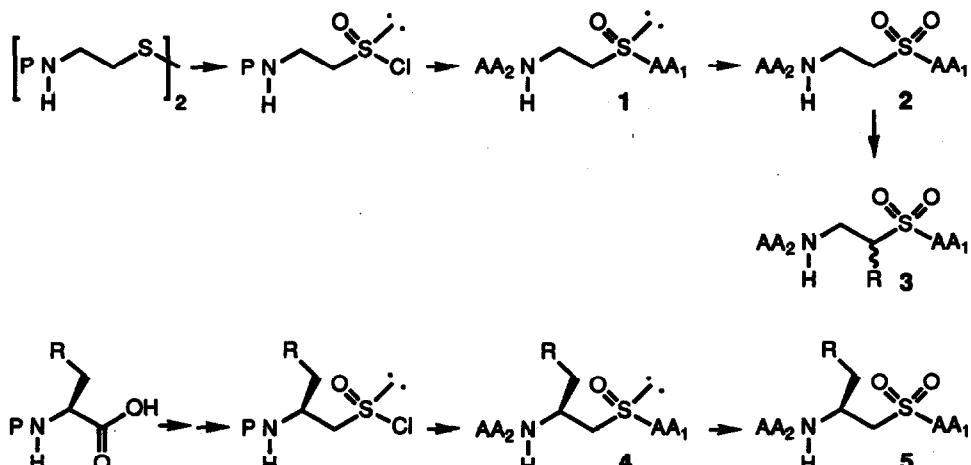
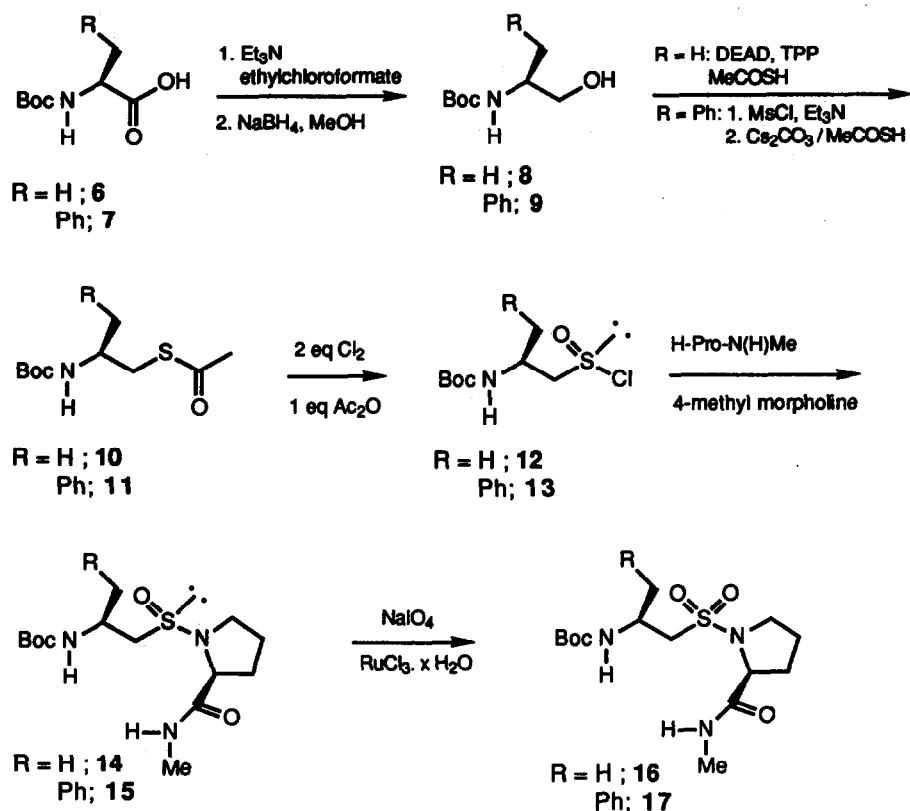


figure 1c

We have illustrated our method with the synthesis of sulfonamide and sulfonamide transition-state isosteres of Ala-Pro and Phe-Pro. Furthermore, as a specific target molecule we chose to synthesize the isostere of Phe-Phe, which will be used as a hapten to generate catalytic antibodies¹¹.



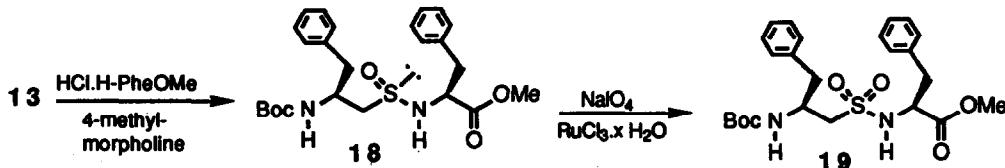
Scheme 1



Scheme 2

The Boc protected aminoacids **6** and **7** were converted to the corresponding amino alcohols **8** and **9** by reduction of the of the *in situ* prepared mixed anhydrides in 68% and 67% yield respectively¹². The alaninol derivative **8** could be converted to the thioester **10** in 85% yield using Mitsunobu conditions^{10,13}. Applying this procedure to phenylalaninol derivative **9** gave the desired product **11** from which, unfortunately, triphenylphosphinoxide could not be separated completely. Therefore **11** was prepared by a two step procedure i.e. formation of the mesylate (96% yield) followed by substitution with cesium thioacetate (97% yield)^{10,13,14}. The sulfinylchlorides **12** and **13** were prepared by treatment of the thioesters **10** and **11** with chlorine (approximately 2 eq.) in the presence of acetic anhydride (1 eq.)^{15,16}. These were used for coupling without further purification. Sulfinylchlorides **12** and **13** were coupled to H-Pro(N)HMe¹⁷ in the presence of 4-methylmorpholine as base leading to the peptide sulfinamides **14** (72%) and **15** (75%), respectively. The diastereomers of **14** and **15** were isolated in a approximately equal amounts. The corresponding sulfonamides **16** and **17** were obtained by oxidation using RuCl₃/NaIO₄¹⁸ in 95% and 96% yield, respectively¹⁹ (scheme 2).

In order to prepare the sulfonamide isostere of Phe-Phe (figure 1c), sulfinylchloride **13** was coupled to H-Phe-OMe in the presence of 4-methylmorpholine as base. The resulting diastereomeric sulfinamides **18** were isolated in 66% yield (diastereomeric ratio 1 to 1.6). Oxidation afforded the sulfonamide isostere **19** in 79% yield¹⁹ (scheme 3).



Scheme 3

In conclusion, we have described a straightforward method for the synthesis of homochiral β -functionalized amineethane sulfinamides and sulfonamides. This method enables us to employ in principle every possible α -amino acid in the preparation of sulfinamide or sulfonamide transition-state analogue containing peptides. These peptides might be important both for the development of protease inhibitors and catalytic antibodies.

Acknowledgement. We wish to thank A.W.M. Lefever for recording the 300 MHz NMR spectra and the Dutch AIDS fund for financial support.

References and notes:

- See e.g. Bartlett, P.A.; Marlowe, C.K. *Science* **1987**, *235*, 569-571; Allen, M.C.; Fuhrer, W.; Tuck, B.; Wade, R.; Wood, J.M. *J. Med. Chem.* **1989**, *32*, 1652-1661; Melnick, M.J.; Bisaha, S.N.; Gammill, R.B. *Tetrahedron Lett.* **1990**, *31*, 961-964; Nakano, M.; Atsuumi, S.; Koike, Y.; Tanaka, S.; Funabashi, H.; Hashimoto, J.; Ohkubo, M.; Morishima, H. *Chem. Lett.* **1990**, 505-508; Dreyer, G.B.; Metcalf, B.W.; Tomaszek Jr., T.A.; Carr, T.J.; Chandler III, A.C.; Hyland, L.; Fakhoury, S.A.; Magaard, V.W.; Moore, M.L.; Strickler, J.E.; Debouck, C.; Meek, T.D. *Proc. Natl. Acad. Sci. U.S.A.* **1989**, *86*, 9752-9756; Roberts, N.A.; Martin, J.A.; Kinchington, D.; Broadhurst, A.V.; Craig, J.C.; Duncan, I.B.; Galpin, S.A.; Handa, B.K.; Kay, J.; Kröhn, A.; Lambert, R.W.; Merrett, J.H.; Mills, J.S.; Parkes, K.E.B.; Redshaw, S.; Ritchie, A.J.; Taylor, D.L.; Thomas, G.J.; Machin, P.J. *Science*, **1990**, *248*, 358-361; Rich, D.H.; Green, J.; Toth, M.V.; Marshall, G.R.; Kent, S.B.H. *J. Med. Chem.* **1990**, *33*, 1285-1288.
- Janda, K.D.; Schloeder, D.; Benkovic, S.J.; Lerner, R.A. *Science* **1988**, *241*, 1188-1191; Iverson, B.L.; Lerner, R.A. *Science* **1989**, *243*, 1184-1188.
- See e.g. Chakravarty, P.K.; de Laszlo, S.E.; Sarnella, C.S.; Springer, J.P.; Schuda, P.F. *Tetrahedron Lett.* **1989**, *30*, 415-418; Mock, W.L.; Tsay, J.-T. *J. Am. Chem. Soc.* **1989**, *111*, 4467-4472

4. Merricks, D.; Sammes, P.G.; Walker, E.R.H.; Henrick, K.; McPartlin, M.M. *J. Chem. Soc. Perkin Trans. I*, 1991, 2169-2176.
5. Calcagni, A.; Gavuzzo, E.; Lucente, G.; Mazza, F.; Pochetti, G.; Rossi, D. *Int. J. Peptide Protein Res.* 1989, 34, 319-324; Calcagni, A.; Gavuzzo, E.; Lucente, G.; Mazza, F.; Pinnen, F.; Pochetti, G.; Rossi, D. *Ibid.* 1989, 34, 471-479; Calcagni, A.; Gavuzzo, E.; Lucente, G.; Mazza, F.; Pinnen, F.; Pochetti, G.; Rossi, D. *Ibid.* 1991, 37, 167-173; Zecchini, G.P.; Paradisi, M.P.; Torrini, I.; Lucente, G.; Gavuzzo, E.; Mazza, F.; Pochetti, G. *Tetrahedron Lett.* 1991, 32, 6779-6782.
6. Moree, W.J.; Van der Marel, G.A.; Liskamp, R.M.J. *Tetrahedron Lett.* 1991, 32, 409-412.
7. Norbeck, D.W. *Ann. Rep. Med. Chem.* 1990, 25, 149.
8. Moree, W.J.; Van Gent, L.C.; Van der Klein-de Gunst, F.J.M.; Van der Marel G.A.; Liskamp, R.M.J. Poster presented at Twelfth American Peptide Symposium, Boston, June 16-21, 1991; Moree, W.J.; Van Gent, L.C.; Van der Marel, G.A.; Liskamp, R.M.J. *manuscript in preparation*
9. Neelakantan, L.; Hartung, W.H. *J. Org. Chem.* 1959, 24, 1943-1948; Frankel, M.; Moses, P. *Tetrahedron* 1960, 9, 289-294; Gilmore, W. F.; Lin, H-J. *J. Org. Chem.* 1978, 43, 4535-4537.
10. Higashiura, K.; Ienaga, K. *J. Org. Chem.* 1992, 57, 764-766.
11. Janda, K.D. (The Scripps Research Institute, La Jolla, California 92037, USA) *Personal communication*.
12. Kokotos, G. *Synthesis* 1990, 299-301.
13. Volante, R.P. *Tetrahedron Lett.* 1981, 22, 3119-3122.
14. Strijtveen, B.; Kellogg, R.M. *J. Org. Chem.* 1986, 51, 3664-3671.
15. Kee, M-L.; Douglass, I.B. *Org. Prep. and Proc.* 1970, 2, 235-244.
16. Van den Broek, L.A.G.M.; Lázaro, E.; Zylicz, Z.; Fennis, P.J.; Missler, F.A.N.; Lelieveld, P.; Garzotto, M.; Wagener, D.J. T.; Ballesta, J.P.G.; Ottenheijm, H.C.J. *J. Med. Chem.* 1989, 32, 2002-2015.
17. H-Pro-N(H)Me was prepared from Z-Proline: formation of the methyl amide by the mixed anhydride method followed by hydrogenolysis of the Z-group.
18. Gao, Y.; Sharpless, K.B.; *J. Am. Chem. Soc.* 1988, 110, 7538-7539.
19. Compounds were characterized by ¹H and ¹³C NMR e.g.
 - 14: ¹³C NMR (CDCl₃): 21.1 (AlaS-C³), 25.3 (Pro-C⁴), 26.1 (N(H)CH₃), 28.2 ((CH₃)₃C), 32.1 (Pro-C³), 42.6 (AlaS-C²), 52.6 (Pro-C⁵), 56.9 (Pro-C²), 61.2 (CH₂SO), 79.7 ((CH₃)C), 155.1 (C=O Boc), 173.2 (C=O) ¹³C NMR (CDCl₃): 21.0 (AlaS-C³), 24.7 (Pro-C⁴), 26.2 (N(H)CH₃), 28.3 ((CH₃)₃C), 31.4 (Pro-C³), 40.7 (Pro-C⁵) 42.5 (AlaS-C²), 61.4 (CH₂SO), 67.1 (Pro-C²), 80.0 ((CH₃)C), 155.2 (C=O Boc), 172.7 (C=O)
 - 16: ¹³C NMR (CDCl₃): 20.4 (AlaS-C³), 24.5 (Pro-C⁴), 26.0 (N(H)CH₃), 28.0 ((CH₃)₃C), 30.6 (Pro-C³), 42.8 (AlaS-C²) 48.9 (Pro-C⁵), 54.2 (CH₂SO₂), 61.4 (Pro-C²), 79.3 ((CH₃)C), 154.9 (C=O Boc), 171.9 (C=O)
 - 17: ¹³C NMR (CDCl₃): 24.7 (Pro-C⁴), 26.2 (N(H)CH₃), 28.2 ((CH₃)₃C), 30.6 (Pro-C³), 39.8 (PheS-C³), 48.6 (PheS-C²), 49.1 (Pro-C⁵), 51.5 (CH₂SO₂), 61.8 (Pro-C²), 79.9 ((CH₃)C), 126.8, 128.6, 129.2, 137.0 (PheS-Ph), 155.1 (C=O Boc), 171.8 (C=O)
 - 19: ¹³C NMR (CDCl₃): 28.2 ((CH₃)₃C), 39.2, 40.5 (Phe-C³, PheS-C³) 47.7 (PheS-C²), 52.4 (OMe), 56.4 (CH₂SO₂), 57.1 (Phe-C²), 80.0 ((CH₃)C), 126.8, 127.2, 128.5, 128.6, 129.4, 135.4, 136.5 (Phe-Ph, PheS-Ph), 154.1 (C=O Boc), 172.0 (C=O)

(Received in UK 9 July 1992)